

SHORT REPORT

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# Mutated $Ca_v2.1$ channels dysregulate CASK/P2X3 signaling in mouse trigeminal sensory neurons of R192Q *Cacna1a* knock-in mice

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## Abstract

**Background:** ATP-gated P2X3 receptors of sensory ganglion neurons are important transducers of pain as they adapt their expression and function in response to acute and chronic nociceptive signals. The present study investigated the role of calcium/calmodulin-dependent serine protein kinase (CASK) in controlling P2X3 receptor expression and function in trigeminal ganglia from *Cacna1a* R192Q-mutated knock-in (KI) mice, a genetic model for familial hemiplegic migraine type-1.

**Results:** KI ganglion neurons showed more abundant CASK/P2X3 receptor complex at membrane level, a result that likely originated from gain-of-function effects of R192Q-mutated  $Ca_v2.1$  channels and downstream enhanced CaMKII activity. The selective  $Ca_v2.1$  channel blocker  $\omega$ -Agatoxin IVA and the CaMKII inhibitor KN-93 were sufficient to return CASK/P2X3 co-expression to WT levels. After CASK silencing, P2X3 receptor expression was decreased in both WT and KI ganglia, supporting the role of CASK in P2X3 receptor stabilization. This process was functionally observed as reduced P2X3 receptor currents.

**Conclusions:** We propose that, in trigeminal sensory neurons, the CASK/P2X3 complex has a dynamic nature depending on intracellular calcium and related signaling, that are enhanced in a transgenic mouse model of genetic hemiplegic migraine.

**Keywords:** Pain, ATP, Purinergic signaling, DRG, Trigeminal ganglion, Migraine

## Background

P2X3 receptors are predominantly expressed on sensory ganglion neurons where they play an important role in transducing pain signals [1]. A major property of these receptors is the ability to rapidly adapt their function to extracellular milieu changes by trafficking-mediated receptor redistribution, by modulation of receptor function through intracellular kinases, or by interaction with specific scaffold proteins [2-5]. We recently reported that under basal conditions P2X3 receptors are strongly associated with the multifunction scaffold protein calcium/calmodulin-dependent serine protein kinase (CASK) [6].

In the present study we investigated whether the CASK/P2X3 complex was altered and functionally linked to sensitization of P2X3 receptors in transgenic knock-in (KI) mice exhibiting a gain-of-function phenotype of voltage-gated  $Ca_v2.1$  (P/Q-type) calcium channels, due to a R192Q missense mutation in the channel  $\alpha1$  subunit that causes familial hemiplegic migraine type 1 (FHM-1) [7,8]. Using this KI mouse model, we previously identified multiple  $Ca_v2.1$  channel interactors (calcineurin, Cdk5 and CaMKII) that modulate P2X3 receptor function in trigeminal sensory neurons [9-12]. In particular, enhanced P2X3 receptor-mediated responses were found in KI neurons that depend on constitutive activation of CaMKII and are reversed by the selective  $Ca_v2.1$  channel blocker or by the CaMKII inhibitor [9]. Previous studies showed that CASK is associated with calcium channels [13-15] and, thus, provide the rationale to explore if the

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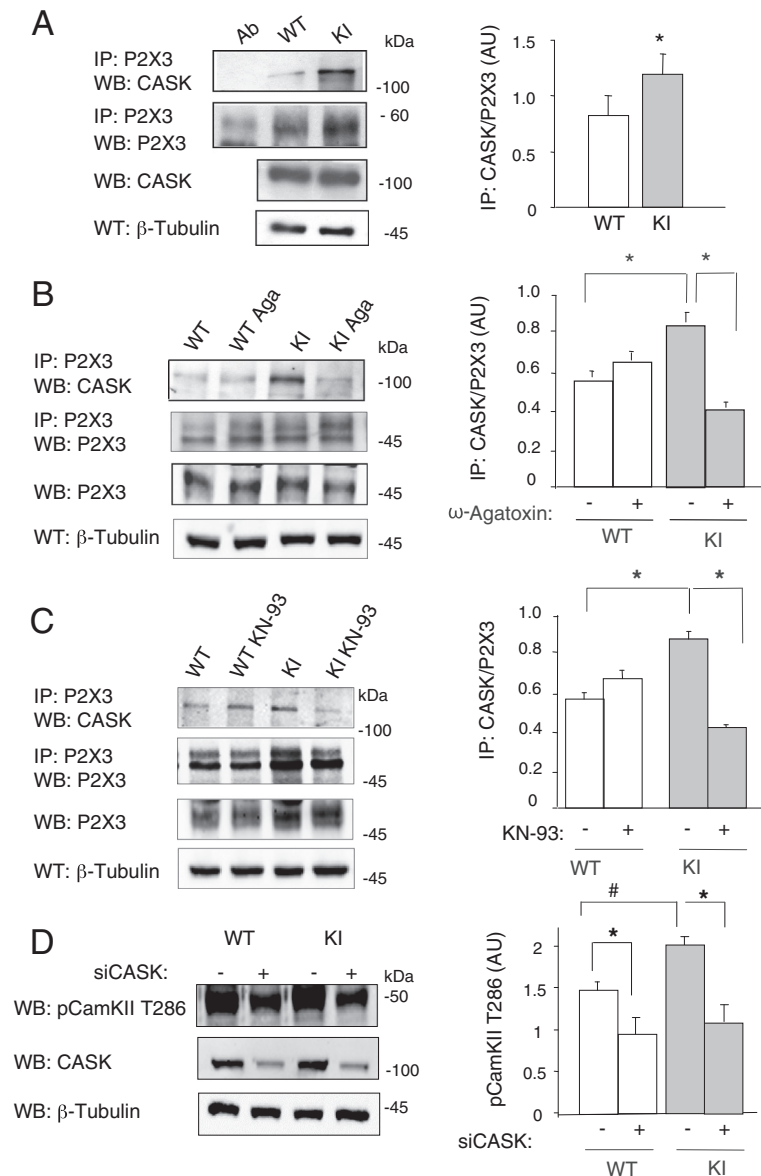
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R192Q mutation in KI mice influences CASK/P2X3 assembly and function. The present study aimed at testing, with molecular biology and electrophysiological methods, the properties of the CASK/P2X3 receptor complex in this mouse model expressing gain-of-function of  $Ca_v2.1$  channels, using primary cultures of trigeminal ganglia that fully retain the basal characteristics of the CASK/P2X3 complex *in vivo* [6].

## Results

### The CASK/P2X3 receptor complex is abundantly expressed in KI ganglia and is modulated by $Ca^{2+}$ influx

In order to study the effects of CASK on P2X3 receptors expressed in WT and KI ganglia, we first compared CASK/P2X3 complex levels in ganglion extracts. Immunoprecipitation experiments showed that the complex was significantly more abundant in KI than in WT samples



**Figure 1 CASK/P2X3 complex in KI trigeminal neurons.** **A**, Example of immunopurified P2X3 from trigeminal ganglia probed with anti-CASK antibodies reveals more abundant CASK/P2X3 complex in KI ganglion cultures than in WT ones. Histograms quantify this effect ( $n = 5$ ,  $p = 0.038$ ). Ab, indicate signal from unrelated antibody. **B**, CASK/P2X3 co-immunoprecipitation experiments from ganglion cultures demonstrate that CASK/P2X3 complex is sensitive to  $\omega$ -Agatoxin IVA treatment (400 nM, overnight; Aga). Note the more intense CASK/P2X3 signal in KI samples as seen in **A**. P2X3 immunoprecipitation input and  $\beta$ -Tubulin expression shown as gel loading controls. **C**, CASK/P2X3 complex is sensitive to KN-93 pre-incubation (5  $\mu$ M, 90 min). **D**, Examples of anti-phospho-CaMKII T286 Western immunoblots of extracts from WT and KI trigeminal cultures in control conditions and after CASK silencing.  $\beta$ -Tubulin is shown as loading control. Histograms quantify the effect (#,  $p = 0.05$ ; \* $p = 0.03$  for WT and  $p = 0.025$  for KI;  $n = 3$ ).

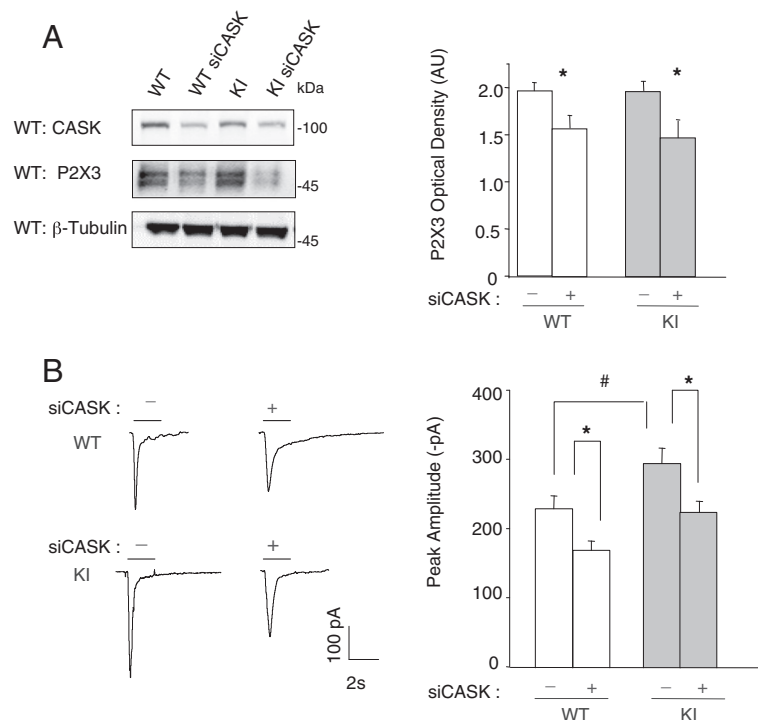
(Figure 1A;  $p = 0.038$ ,  $n = 5$ ). A significant increase ( $n = 5$ ,  $p = 0.005$ ) in CASK associated with cell membrane fractions was observed in KI tissue (Additional file 1: Figure S1A), although total CASK lysate preparations did not show any difference between WT or KI samples (Additional file 1: Figure S1B). Further experiments concerning the specificity of the CASK/P2X3 complex, based on immunoprecipitating CASK first and then performing western blotting with P2X3 antibodies, validated our previous findings [6] and are included in Additional file 2: Figure S2A, B.

In analogy to its effect on other receptors (e.g. NMDA receptors; [15]), CASK might exert a role in the process of P2X3 receptor export to surface membranes. In fact, pulled-down biotinylated surface P2X3 receptors showed co-purification with intracellular CASK (Additional file 3: Figure S3), supporting the view that CASK/P2X3 complexes are membrane-bound. In these biotinylation experiments, no difference was observed in the levels of surface membrane CASK in WT and KI samples (Additional file 3: Figure S3).

We further explored whether the origin of the stronger CASK/P2X3 association in KI samples could be

directly linked to the  $Ca_v2.1$  R192Q gain-of-function and concomitant higher CaMKII activation [9]. To this end, we used selective pharmacological tools to specifically block either  $Ca_v2.1$  channel function or apply inhibitors of downstream CaMKII activity. Trigeminal ganglion cultures were treated with the selective  $Ca_v2.1$  channel blocker  $\omega$ -Agatoxin IVA (400 nM, 15 h) and then tested for CASK/P2X3 protein interaction by immunoprecipitation. Immunoprecipitation experiments with purified P2X3 receptors and CASK expression (Figure 1B) confirmed that CASK/P2X3 receptor interaction was stronger in KI cultures and that it was significantly reversed by  $\omega$ -Agatoxin IVA application. Interestingly, CASK/P2X3 complex in WT culture was not affected by the channel blocker suggesting that larger CASK/P2X3 interaction in KI neurons was likely due to the enhanced  $Ca_v2.1$  channel activity.

Likewise, after cultures treatment with the CaMKII inhibitor KN-93 (5  $\mu$ M, 90 min, Figure 1C), the CASK/P2X3 interaction was significantly inhibited in KI, but not in WT (Figure 1C). In addition, after RNA silencing of CASK expression [6], there was a significant decrease in CaMKII phosphorylation (Figure 1D).



**Figure 2 Effect of siCASK silencing in KI and WT neurons.** **A**, Example of Western immunoblotting for CASK, P2X3 and  $\beta$ -tubulin, in control and after siCASK silencing. Note the significant decrease in P2X3 receptor expression in silenced samples ( $*p = 0.026$  for WT and  $p = 0.04$  for KI;  $n = 5$ , number of experiments). **B**, representative examples of currents induced by  $\alpha, \beta$ -meATP ( $\alpha, \beta$ , black bar, 10  $\mu$ M) from WT or KI trigeminal neurons in control or after siCASK. Histograms show quantification of mean current amplitude values (WT:  $230 \pm 19$  pA or  $170 \pm 13$  pA; KI:  $296 \pm 92$  pA or  $225 \pm 16$  pA for control or siCASK respectively). #  $p = 0.036$ ,  $*p = 0.02$  for WT and  $p = 0.019$  for KI;  $n = 15$ –23 cells.

### P2X3 expression and function after siCASK in WT and KI ganglion cultures

Our recent findings [6] that showed how siCASK significantly lowered P2X3 expression in trigeminal ganglion cultures, have been further validated in the present study in which no difference between WT and KI cultures was observed as a consequence of siCASK (Figure 2A). To further explore functional consequence of CASK/P2X3 complex in the KI model, patch clamp experiments were carried out (Figure 2B). Sample P2X3 receptor currents elicited by pulse application of the selective agonist  $\alpha,\beta$ -methylene-adenosine-5'-triphosphate ( $\alpha,\beta$ -meATP; 10  $\mu$ M) were clearly smaller after siCASK silencing, but proportionally similar in WT and KI neurons. As expected [9], KI neuronal currents were constitutively larger than WT ones (Figure 2B).

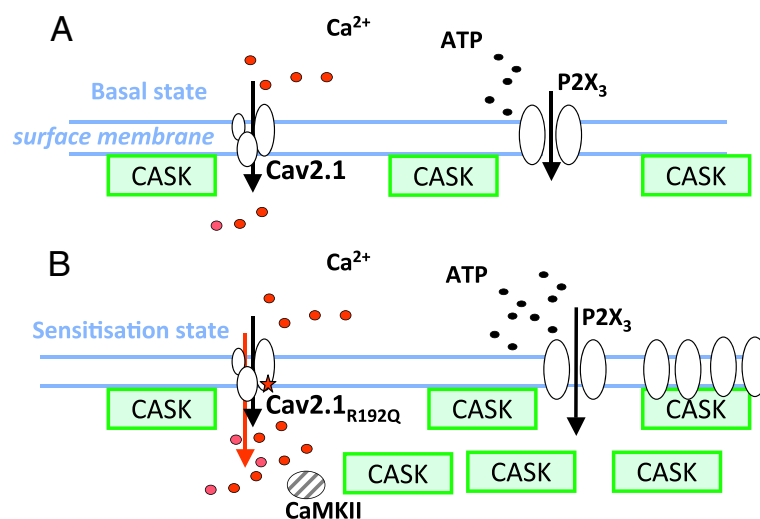
### Discussion

The principal finding of the present study is the demonstration that the R192Q missense mutation in the  $\alpha 1$  subunit of  $Ca_v2.1$  calcium channels, that confers gain of function to these channels, is associated with more abundant CASK/P2X3 complex in trigeminal sensory neurons with a mechanism clearly dependent on  $Ca_v2.1$  channel function and CaMKII activation. This result outlines a molecular mechanism whereby P2X3 receptors are retained and display stronger activity in KI neurons and can, therefore, contribute to sensitization to painful stimuli.

Our former studies have shown that P2X3 receptors of trigeminal sensory neurons are under the influence of endogenous kinases such as Csk and Cdk5, which

regulate the basal operational activity of these receptors [9,16]. We have recently observed a new member of this class of regulators, namely CASK, which strongly controls P2X3 receptor expression and function at the plasma membrane of mouse trigeminal neurons under physiological conditions [6,17]. The present report shows that CASK is more associated with P2X3 receptors in KI ganglia and in culture, and that this association is largely dependent on  $Ca_v2.1$  calcium channel and CaMKII activity as their specific pharmacological blockers reversed the effect. When CASK expression was inhibited by siRNA silencing, the P2X3 receptor expression and function fell to similar levels in WT and KI neurons. In biotinylation experiments, since no difference was observed in CASK fractions from WT or KI surface membrane samples, these experiments are, therefore, consistent with a differential distribution of P2X3 receptors to lipid raft compartments in KI ganglia [3]. When CASK was blocked, the residual P2X3 receptor currents were approximately one-third smaller, providing a first estimate about the extent of the role of CASK in regulating P2X3 receptor activity. Such data suggest that the stronger CASK/P2X3 association in KI cells was an important factor for the observed up-regulation of P2X3 receptors.

Figure 3 summarizes an idealized scheme of how we envisage the interaction between certain neuronal proteins affecting P2X3 receptor responsiveness. In WT neurons, only a fraction of P2X3 receptors is thought to be associated with CASK, which ensures their stability at



**Figure 3 Proposed mechanism of action of CASK/P2X3 complex in trigeminal sensory neurons.** Schematic representation of CASK, P2X3, and  $Ca_v2.1$  channel protein interactions at the plasma membrane of trigeminal sensory neurons. **A**, In WT ganglion cells, only a fraction of P2X3 receptors is thought to be associated with CASK that ensures their stable expression. Under these conditions,  $Ca_v2.1$  channels provide only a minor contribution to  $Ca^{2+}$  influx, and the dynamic CASK/P2X3 association ensures physiological responses and P2X3 receptors turn-over [6]. **B**, In KI ganglia, the hyperfunctional  $Ca_v2.1$  channels result in a stronger  $Ca^{2+}$  influx, larger CaMKII activity, and more abundant CASK/P2X3 receptor complexes with improved stability of P2X3 receptors. Sub-membrane vesicle-associated CASK, potentially associated with P2X3 receptor trafficking, is also indicated. Larger release of extracellular ATP [20] facilitates P2X3 responses and may contribute to the process of sensitization of P2X3 receptors.

the plasma membrane level [6]. Under these conditions, Ca<sub>v</sub>2.1 channel activity provides only a relatively minor contribution to Ca<sup>2+</sup> influx [9,18,19] and P2X3 receptors are regulated only to limited extent by CASK (Csk and Cdk5 are not depicted here). The hyper-functional Ca<sub>v</sub>2.1 channels in KI (that require a smaller membrane depolarization to reach the activation threshold [8,20]) induce stronger Ca<sup>2+</sup> influx and CaMKII activity [9], improving the stability of CASK/P2X3 receptor complexes and their activity at plasma membrane level. Furthermore, these processes probably synergize and add to the already larger release of extracellular ATP by KI ganglia [21], thereby further facilitating P2X3 responses and thus contributing to the process of sensitization of P2X3 receptors [9].

In conclusion, the present data suggest that CASK could play the role of scaffold protein linking different membrane and sub-membrane molecules (including other channels) to elicit further downstream signaling. Since CASK is involved at presynaptic level to modulate synaptic transmitter release [22], it is feasible that this anchoring mechanism may also occur at central level and shows distinct alterations in chronic pain.

## Materials and methods

### Tissue culture

Ca<sub>v</sub>2.1 R192Q KI and WT mice [8] were euthanized by slowly raising levels of CO<sub>2</sub> in accordance with the Italian Animal Welfare Act and approved by SISSA Ethical Committee. Trigeminal ganglia primary cultures were obtained as previously described [6]. In specific experiments ω-Agatoxin IVA (400 nM; 15 h; Sigma, Milan, Italy) or CaMKII inhibitor KN-93 (5 μM; Sigma) were added to the culture medium for the indicated time [9]. CASK siRNA experiments were performed as previously reported [6].

### Western blotting and immunoprecipitation

Protein extracts and immunoprecipitation experiments and membrane protein biotinylation were performed as previously described [6]. P2X3 receptors were solubilized in 10 mM Tris-HCl (pH 7.5), 150 mM NaCl, 20 mM EDTA, and 1% Triton-X-100 plus protease inhibitors (Roche Products, Welwyn Garden City, UK) as previously reported [3,6]. The non-ionic detergent n-octyl-β-D-glucoside (ODG; [9]) was not used. For western blot quantification, signals expressed in optical density absolute units (AU) were measured with digital imaging system (UVITEC, Cambridge, UK).

### Patch clamp recordings

Currents from siRNA-treated cultures (72 h) were recorded under whole cell voltage clamp mode at holding potential of -65 mV after correcting for the liquid

junction potential [6]. P2X3 receptor synthetic agonist α,β-meATP (10 μM; Sigma) was applied (2 s) using a fast superfusion system (RSC-200; BioLogic Science Instruments, Claix, France).

### Data analysis

Data are presented as mean ± standard error. Statistical significance was evaluated with paired Student's *t*-test or Mann-Whitney rank sum test using OriginPro 7.5 (OriginLab, Northampton, MA, USA).

### Additional files

**Additional file 1: Figure S1. A, B** Example of a Western blot experiment of total trigeminal ganglia extracts (**A**) or total membrane or ganglia samples (**B**) from WT and KI mice tested with anti-CASK antibodies. Histogram quantifications show no difference of total CASK expression in WT and KI (*n* = 3; *p* > 0.05), and a significant enrichment of total membrane-associated CASK expression in KI vs WT (*n* = 5, *p* = 0.005). Actin was used as gel loading control.

**Additional file 2: Figure S2. A**, Western blot experiment of trigeminal ganglia extracts after immunoprecipitation with anti-CASK and revealed with anti-CASK or anti-P2X3 antibodies. β-Actin signals quantify immunoprecipitation input. **B**, Membrane protein biotinylation experiments of trigeminal ganglia cultures in control and after siP2X3, analysed with western blot and probed with anti-CASK or anti-P2X3 antibodies, as indicated. Signals from total extracts (left) and streptavidin pull-down (right) of biotinylated samples are shown. No difference in membrane-bound CASK after siP2X3 is found β-Actin is used as gel loading control. Note lack of β-Actin in pull-down samples.

**Additional file 3: Figure S3**. Membrane protein biotinylation experiments of WT and KI trigeminal ganglia cultures in control and after siP2X3, revealed with western blot and probed with anti-CASK and anti-P2X3 antibodies. Signals from total extracts (left) and streptavidin pull-down (right) of biotinylated samples are shown. β-Actin is used as gel loading control. Histograms quantify larger surface P2X3 receptors in trigeminal ganglia cultures from KI mice and no changes in surface-associated CASK between WT and KI samples (*n* = 3, \**p* < 0.05).

### Abbreviations

α: β-meATP α,β-methyleneATP; CaMK: Calcium/calmodulin-dependent kinase; CASK: Calcium/calmodulin-dependent serine kinase; TG: Trigeminal ganglia.

### Competing interests

The authors state that the content of this article does not create any competing interest.

### Authors' contributions

AG carried out biochemical studies and statistical analysis, TB performed further data collection, SH carried out functional studies and statistical analysis, MF, AM provided animal model; EF, MS conceived of the study, EF, AN design and coordination; AG, AM, AN, EF draft the manuscript. All authors read and approved the final manuscript.

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## References

1. Burnstock G: **Purinergic signalling and disorders of the central nervous system.** *Nat Rev Drug Discov* 2008, **7**:575–590.
2. Chen XQ, Wang B, Wu C, Pan J, Yuan B, Su YY, Jiang XY, Zhang X, Bao L: **Endosome-mediated retrograde axonal transport of P2X3 receptor signals in primary sensory neurons.** *Cell Res* 2012, **22**:677–696.
3. Gnanasekaran A, Sundukova M, van den Maagdenberg AM, Fabbretti E, Nistri A: **Lipid rafts control P2X3 receptor distribution and function in trigeminal sensory neurons of a transgenic migraine mouse model.** *Mol Pain* 2011, **7**:77.
4. Vacca F, D'Ambrosi N, Nestola V, Amadio S, Giustizieri M, Cucchiaroni ML, Tozzi A, Velluz MC, Mercuri NB, Volonté C: **N-Glycans mutations rule oligomeric assembly and functional expression of P2X3 receptor for extracellular ATP.** *Glycobiology* 2011, **21**:634–643.
5. Vacca F, Giustizieri M, Ciotti MT, Mercuri NB, Volonté C: **Rapid constitutive and ligand-activated endocytic trafficking of P2X receptor.** *J Neurochem* 2009, **109**:1031–1041.
6. Gnanasekaran A, Sundukova M, Hullugundi S, Birsa N, Bianchini G, Hsueh YP, Nistri A, Fabbretti E: **CASK is a new intracellular modulator of P2X3 receptors.** *J Neurochem* 2013, **126**:102–112.
7. Ophoff RA, Terwindt GM, Vergouwe MN, van Eijk R, Oefner PJ, Hoffman SM, Lamerdin JE, Mohrenweiser HW, Bulman DE, Ferrari M, Haan J, Lindhout D, van Ommen GJ, Hofker MH, Ferrari MD, Frants RR: **Familial hemiplegic migraine and episodic ataxia type-2 are caused by mutations in the Ca<sup>2+</sup> channel gene CACNL1A4.** *Cell* 1996, **87**:543–552.
8. van den Maagdenberg AM, Pietrobon D, Pizzorusso T, Kaja S, Broos LA, Cesetti T, van de Ven RC, Tottene A, van der Kaa J, Plomp JJ, Frants RR, Ferrari MD: **A Cacna1a knockin migraine mouse model with increased susceptibility to cortical spreading depression.** *Neuron* 2004, **41**:701–710.
9. Nair A, Simonetti M, Birsa N, Ferrari MD, van den Maagdenberg AM, Giniatullin R, Nistri A, Fabbretti E: **Familial hemiplegic migraine Ca<sub>v</sub>2.1 channel mutation R192Q enhances ATP-gated P2X3 receptor activity of mouse sensory ganglion neurons mediating trigeminal pain.** *Mol Pain* 2010, **6**:48.
10. Nair A, Simonetti M, Fabbretti E, Nistri A: **The Cdk5 kinase downregulates ATP-gated ionotropic P2X3 receptor function via serine phosphorylation.** *Cell Mol Neurobiol* 2010, **30**:505–509.
11. Simonetti M, Giniatullin R, Fabbretti E: **Mechanisms mediating the enhanced gene transcription of P2X3 receptor by calcitonin gene-related peptide in trigeminal sensory neurons.** *J Biol Chem* 2008, **283**:18743–18752.
12. Fabbretti E, Nistri A: **Regulation of P2X3 receptor structure and function.** *CNS Neurol Disord Drug Targets* 2012, **11**:687–698.
13. Maximov A, Südhof TC, Bezprozvanny I: **Association of neuronal calcium channels with modular adaptor proteins.** *J Biol Chem* 1999, **274**:24453–24456.
14. Spafford JD, Zamponi GW: **Functional interactions between presynaptic calcium channels and the neurotransmitter release machinery.** *Curr Opin Neurobiol* 2003, **13**:308–314.
15. Jeyifous O, Waites CL, Specht CG, Fujisawa S, Schubert M, Lin EI, Marshall J, Aoki C, de Silva T, Montgomery JM, Garner CC, Green WN: **SAP97 and CASK mediate sorting of NMDA receptors through a previously unknown secretory pathway.** *Nat Neurosci* 2009, **12**:1011–1019.
16. D'Arco M, Giniatullin R, Leone V, Carloni P, Birsa N, Nair A, Nistri A, Fabbretti E: **The C-terminal Src inhibitory kinase (Csk)-mediated tyrosine phosphorylation is a novel molecular mechanism to limit P2X3 receptor function in mouse sensory neurons.** *J Biol Chem* 2009, **284**:21393–21401.
17. Volonté C, Burnstock G: **P2X3 receptor: a novel 'CASKade' of signaling?** *J Neurochem* 2013, **126**:1–3.
18. Ikeda M, Matsumoto S: **Classification of voltage-dependent Ca<sup>2+</sup> channels in trigeminal ganglion neurons from neonatal rats.** *Life Sci* 2003, **73**:1175–1187.
19. Tao J, Liu P, Xiao Z, Zhao H, Gerber BR, Cao YQ: **Effects of familial hemiplegic migraine type 1 mutation T666M on voltage-gated calcium channel activities in trigeminal ganglion neurons.** *J Neurophysiol* 2012, **107**:1666–1680.
20. Tottene A, Conti R, Fabbro A, Vecchia D, Shapovalova M, Santello M, van den Maagdenberg AM, Ferrari MD, Pietrobon D: **Enhanced excitatory transmission at cortical synapses as the basis for facilitated spreading depression in Ca<sub>v</sub>2.1 knockin migraine mice.** *Neuron* 2009, **61**:762–773.
21. Franceschini A, Nair A, Bele T, van den Maagdenberg AM, Nistri A, Fabbretti E: **Functional crosstalk in culture between macrophages and trigeminal sensory neurons of a mouse genetic model of migraine.** *BMC Neurosci* 2012, **13**:143.
22. Hsueh YP: **The role of the MAGUK protein CASK in neural development and synaptic function.** *Curr Med Chem* 2006, **13**:1915–1927.

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